IgA dominant glomerulonephritis associated to staphylococcus infection: a peculiar case report

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ABSTRACT

IgA dominant glomerulonephritis associated to Staphylococcus infection is a rare clinical entity that has been described mainly in case reports. Biopsy features can resemble other disease entities mainly IgA nephropathy and Henoch-Schönlein purpura nephritis. Treatment of IgA dominant glomerulonephritis associated to staphylococcal infection is based on antibiotics for the underlying infection, controlling hypertension and edema and may resort to concomitant use of steroids in selected cases. Prognosis markers such as hypertension, diabetes and interstitial fibrosis may influence treatment as they are associated with poor renal outcomes. We report a case of a 63-year-old man with known hypertension, pre-diabetes and recent history of methicillin-sensitive Staphylococcus aureus (MSSA) bacteremia associated to prostatitis, who presented with a one-month history of edema, arthralgia and foamy urine. Over this period he progressed to anasarca and nephrotic range proteinuria with concomitant rise in creatinine levels being documented. The renal biopsy showed segmental endocapillary proliferation and IgA segmental dominant staining associated to C3 and lambda in minor distribution. On completion of two months of steroid therapy the patient partially recovered his renal function and proteinuria. After nine months of tapering steroids, he presented with acute inflammatory arthritis supporting an inflammatory background disease. To our knowledge this case describes an unusual entity such as IgA dominant glomerulonephritis associated to staphylococcal infection co-presenting with an associated reactive arthritis.

Keywords: IgA nephropathy, Staphylococcus, Postinfectious glomerulonephritis

INTRODUCTION

IgA dominant glomerulonephritis (GN) associated to staphylococcal infection while rare is an increasingly recognized morphologic variant of infectious GN, in which there is glomerular deposition of IgA, either dominant or co-dominant. This entity has been described mainly through minor case reports.1,2 Koyama et al first described it in 1995, in 10 patients with methicillin-resistant Staphylococcus aureus (MRSA) infection.2 Methicillin-sensitive Staphylococcus aureus (MSSA) has also been reported in 18% of patients.2

It is thought that bacterial superantigens, such as enterotoxins (C and A) and toxic shock syndrome toxin act as activators of the immune system, leading to the formation of immunocomplexes.3 By binding to major histocompatibility class II molecules they stimulate T cells and cytokine production, inducing polyclonal activation of IgG and IgA.2 Koyama et al also suggested that a probable adhesion factor, defined as Staphylococcus aureus cell envelope antigen, has a pathogenic induction role since it has been noticed in patients with IgA nephropathy (IgAN) and IgA dominant GN associated to staphylococcal infection but not in other immunocomplex glomerulonephritis.4

IgA dominant GN associated to staphylococcal infection is most frequently seen in elderly male patients and over half are diabetic.5 Staphylococcus has become 3 times more common than Streptococcus as the causative agent of infectious glomerulonephritis in the elderly.6

The origin of infection is highly variable, with the most frequent skin, deep-seated abscesses, surgical wounds, lung, joint and heart.1,5 Features backing IgA dominant GN associated to staphylococcal infection include initial presentation at older age, diabetes, acute renal failure, intercurrent culture documenting staphylococcal infection, hypocomplementemia, diffuse glomerular endocapillary hypercellularity on light microscopy (LM), dominant immunofluorescent (IF) staining for C3 over IgA and presence of subepithelial humps on electron microscopy (EM).7,8 Immunocomplex mediated glomerulonephritis and IgAN have been described as rare entities associated to spondyloarthropathies. Concurrently reactive arthritis and glomerulonephritis are rare among the adult population and often under-diagnosed. Different infectious agents can trigger reactive arthritis, usually affecting people in their second to fourth decades of life and occurs weeks following genitourinary (male to female 9:1) or enteric (1:1) infections.9 Most of these patients respond to non-steroid anti-inflammatory drugs while steroids are effective in peripheral arthritis and aggressive presentation cases.

Regarding the emerging importance of a new entity such as IgA dominant GN associated to Staphylococcus aureus infection we present a patient who had an overlooked MSSA infection history with rapidly
progressive renal dysfunction associated to a classical histological pattern of IgA dominance.

**CASE REPORT**

A 63-year-old caucasian man was referred to our emergency department due to routine laboratory finding of a stage II acute kidney injury (AKI)\(^ {10} \). His medical background was marked by a history of hypertension, diagnosis of pre-diabetes 8 years ago and a recent hospital admission dating back 3 months with a MSSA bacteremia due to prostatitis. At that time, he presented normal kidney function and no proteinuria. His history was otherwise unremarkable as he had no relevant family history and was currently on no chronic medication.

One month before emergency admission the patient referred a prior eight-kilogram weight gain and starting of progressive peripheral edemas, foamy urine and polyarticular pain in small and medium joints with an inflammatory pattern. Apart from the edema, his physical examination was unremarkable, namely for afebrile and normotensive presentations and no skin rash was noticed. Laboratory findings had already shown altered renal function with serum creatinine (sCr) of 1.8mg/dL, hypercholesterolemia and hypertriglyceridemia. Urinalysis was normal, with no proteinuria or hematuria. A laboratory re-evaluation was requested within a one-month period that evidenced an abrupt rise in sCr motivating emergency department referral.

On admission to the emergency department, laboratory tests evidenced a rapidly progressive renal failure, with sCr of 3.62mg/dL as opposed to a 3-months earlier basal level of 0.8mg/dL and 1.8mg/dL one month before. As for urinalysis, an active sediment was now evident showing 3+ erythrocyturia with a nephrotic range proteinuria (4+ with a urinary protein/creatinine ratio of 23mg/mg) and a serum albumin of 22 mg/dL. Subsequent investigation showed no abnormal diagnostic imaging findings on renal ultrasound and a negative serology for viral hepatitis, HIV and VDRL. Immunological markers (anti-nuclear antibody (ANA) (IIF (Indirect Immunofluorescence) and CLIA (chemiluminescent immunoassay), anti-double-stranded DNA (anti-dsDNA) (IFI and CLIA), antineutrophil cytoplasmic antibody (ANCA) (CLIA) were normal. No complement changes were detected and his serum protein electrophoresis showed low albumin, elevated alpha 2 and beta region and decreased gamma region, resembling a nephrotic pattern. Cryoglobulinemia was not screened. Diabetic retinopathy was excluded to assess microvascular diabetic involvement. At this time salt restriction, inhibitor enzyme angiotensin-converting, loop diuretics, statin and low molecular weight heparin (LMWH) were initiated followed by a kidney biopsy on day 3 (with temporary suspension of LMWH) due to a clinical course of rapidly progressive glomerulonephritis. The biopsy showed segmental endocapillary proliferation (Figure 1) and fibrinoid necrosis. Extensive tubular necrosis was noted. IF revealed dominant IgA segmental deposition (Figure 2), associated to C3 and lambda in minor distribution. Kappa light chain was absent. EM was not performed. This histopathological finding associated to his clinical background of diabetes, documented prior 10-week infection of MSSA bacteremia and an aggressive presentation led to the suspicion of IgA dominant GN associated to staphylococcal infection.

Due to his highly active inflammatory histopathological background and no signs of active infection, we opted for a trial of 1mg/kg/day prednisone (60mg) for 8 weeks, then tapered off over a 9-month period. Gradual improvement of his renal function was documented, and after 52 weeks his sCr was 1.8mg/dL (maximum sCr level 4.84mg/dL) (Table I). Though, following steroids withdrawal and apparent renal recovery with no signs of clinical activity (lower level of hematuria and total remission of proteinuria and normal complementemia), the arthralgia relapsed following the same previous pattern but with a

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**Figure 1**

Glomerulus showing mesangial and endocapillary proliferation and infiltrating leucocytes. Periodic acid-Schiff, ×400

**Figure 2**

Anti-IgA immunofluorescence showing granular glomerular capillary wall and mesangial staining for IgA
higher pain intensity (numeric pain scale 7/10) and highly debilitating, raising the suspicion of post infectious arthritis. The patient responded only after the reintroduction of low dose prednisone and hydroxychloroquine.

**DISCUSSION**

This case highlights the often misdiagnosed IgA dominant GN associated to staphylococcal infection that mimics IgAN mainly in cases where infection is either overlooked or not apparent. In this way, IgA dominant GN associated to staphylococcus infection represents a challenge in our clinical practice.

Our patient’s clinical, laboratorial and histological presentation follows literature findings. In some of the major studies, the diagnosis is made if at least two of the following criteria is satisfied in a patient with clinical or laboratory evidence of staphylococcal infection and onset of glomerulonephritis usually with endocapillary proliferation and exudative glomerulonephritis on LM (observed in our patient), C3 dominant/co-dominant glomerular staining or IgA-dominant or co-dominant disease together with intense C3 staining on IF microscopy (observed in our patient) and hump-shaped subepithelial deposits on EM, paired with hypocomplementemia (primarily C3).

Multiple features support IgA dominant GN over IgAN namely intercurrent culture documented staphylococcal infection, older age, history of diabetes mellitus and acute renal failure. The majority of patients present with AKI with concomitant microscopic hematuria in 83% and nephrotic range in 50% of cases. As in this case, infection was not apparent at the time of onset of the glomerulonephritis, as is seen in 25% of the cases described in literature. Although staphylococcal infection can be consistent with renal disease, glomerulonephritis can occur within a ten week period, normally between one to sixteen weeks. Pathological features such as endocapillary proliferation on LM and IgA co dominant deposition in IF also sustain this hypothesis. Endocapillary proliferative and exudative GN are reported in 63% of the cases. IF analysis shows a dominant granular IgA glomerular staining with or without a staining for IgG and/or IgM. Staining for C3 is also seen in the majority of cases (normally stronger than IgA). EM was not performed in our patient; nevertheless a low frequency of subepithelial humps compared with poststreptococcal glomerulonephritis has been described particularly in cases in which infection is subtle or not clinically present. Although hypocomplementemia may be an important clue for diagnosis and is present in the majority of cases, complement levels were not consistently depressed in all case reports.

It is extremely unlikely in this case that IgA dominant GN represents an exacerbation of a pre-existing primary IgAN as most patients are older, have no prior history of glomerulonephritis or proliferative histological findings.

**Table I**

<table>
<thead>
<tr>
<th>Variable</th>
<th>Reference range</th>
<th>1 month before hospital admission</th>
<th>On hospital admission</th>
<th>Following 1 week investigation</th>
<th>Day 5 Prednisolone (60mg)</th>
<th>Day 10 Prednisolone (60mg)</th>
<th>2 months Prednisolone (50mg)</th>
<th>4 months Prednisolone (30mg)</th>
<th>9 months Prednisolone (5mg)</th>
<th>12 months Prednisolone (0mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hemoglobin (g/dl)</td>
<td>13.0-18.0</td>
<td>13.4</td>
<td>13.9</td>
<td>12.2</td>
<td>12.4</td>
<td>12.4</td>
<td>9.4</td>
<td>9.2</td>
<td>10.4</td>
<td>10.2</td>
</tr>
<tr>
<td>White-cell count (per mm³)</td>
<td>4500-11,000</td>
<td>7,700</td>
<td>9,900</td>
<td>7,900</td>
<td>14,600</td>
<td>24,300</td>
<td>15,700</td>
<td>11,800</td>
<td>10,100</td>
<td>12,300</td>
</tr>
<tr>
<td>Platelet count (per mm³)</td>
<td>150,000–450,000</td>
<td>269,000</td>
<td>239,000</td>
<td>302,000</td>
<td>291,000</td>
<td>249,000</td>
<td>178,000</td>
<td>184,000</td>
<td>232,000</td>
<td>272,000</td>
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<tr>
<td>Erythrocyte-sedimentation rate (mm/hr)</td>
<td>0-20</td>
<td>–</td>
<td>–</td>
<td>80</td>
<td>59</td>
<td>52</td>
<td>54</td>
<td>35</td>
<td>69</td>
<td>74</td>
</tr>
<tr>
<td>C-reactive protein (mg/dL)</td>
<td>&lt;5.10</td>
<td>–</td>
<td>0-21</td>
<td>2.76</td>
<td>0.61</td>
<td>0.69</td>
<td>0.44</td>
<td>0.40</td>
<td>6.66</td>
<td>16.91</td>
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<td>Urea nitrogen (mg/dl)</td>
<td>8.0-50.0</td>
<td>45</td>
<td>73</td>
<td>55</td>
<td>207</td>
<td>137</td>
<td>139.9</td>
<td>174.7</td>
<td>150.2</td>
<td>129.0</td>
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<tr>
<td>Creatinine (mg/dl)</td>
<td>0.7-1.20</td>
<td>1.8</td>
<td>3.62</td>
<td>3.4</td>
<td>4.84</td>
<td>2.96</td>
<td>1.87</td>
<td>2.07</td>
<td>2.02</td>
<td>1.86</td>
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<tr>
<td>Total protein (g/dl)</td>
<td>60.0-80.0</td>
<td>46.4</td>
<td>46.2</td>
<td>–</td>
<td>41.2</td>
<td>38.4</td>
<td>42.8</td>
<td>48.7</td>
<td>60.3</td>
<td>61.7</td>
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<tr>
<td>Albumin (mg/dl)</td>
<td>35.0-48.0</td>
<td>22</td>
<td>22</td>
<td>19</td>
<td>21</td>
<td>21</td>
<td>25.8</td>
<td>29.9</td>
<td>39.3</td>
<td>40</td>
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<tr>
<td>Total Cholesterol (mg/dl)</td>
<td>&lt;200</td>
<td>–</td>
<td>–</td>
<td>431</td>
<td>–</td>
<td>–</td>
<td>338</td>
<td>215</td>
<td>180</td>
<td>147</td>
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<tr>
<td>Triglyceride (mg/dl)</td>
<td>&lt;150</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>717</td>
<td>367</td>
<td>332</td>
<td>393</td>
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<tr>
<td>C3</td>
<td>90.0-180.0</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>142</td>
<td>120</td>
<td>–</td>
<td>104</td>
<td>142</td>
<td>–</td>
</tr>
<tr>
<td>Protein</td>
<td>negative</td>
<td>negative</td>
<td>4+</td>
<td>1000 mg/dl</td>
<td>400 mg/dl</td>
<td>400 mg/dl</td>
<td>1000 mg/dl</td>
<td>–</td>
<td>100 mg/dl</td>
<td>–</td>
</tr>
<tr>
<td>Red cells</td>
<td>negative</td>
<td>negative</td>
<td>3+</td>
<td>114.3/µl</td>
<td>45.47/µl</td>
<td>291.41/µl</td>
<td>45.47/µl</td>
<td>–</td>
<td>45.47/µl</td>
<td>6.11/µl</td>
</tr>
<tr>
<td>Leukocytes</td>
<td>negative</td>
<td>negative</td>
<td>0-2</td>
<td>12.3/µl</td>
<td>4.2/µl</td>
<td>4.01/µl</td>
<td>0/µl</td>
<td>–</td>
<td>25.7/µl</td>
<td>3.3/µl</td>
</tr>
<tr>
<td>Ratio of total protein (in mg) to creatinine (in mg)</td>
<td>&lt;0.2</td>
<td>negative</td>
<td>–</td>
<td>23.3</td>
<td>13</td>
<td>17.3</td>
<td>10.1</td>
<td>3.6</td>
<td>0.68</td>
<td>0.4</td>
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</tbody>
</table>
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References


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